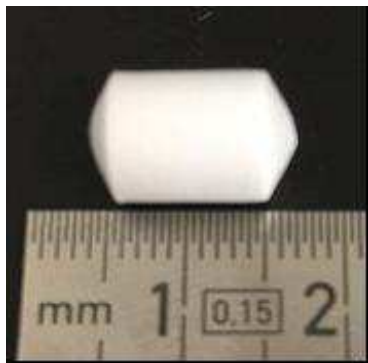


Protocol to make a cellnest sponge



1. The sponge inner structure and shape can be controlled by the concentration of cellnest in the water solution, freezing conditions and mold.
2. *In vivo* degradability can be controlled by crosslinking temperature and time.

[Preparation]

<Materials>

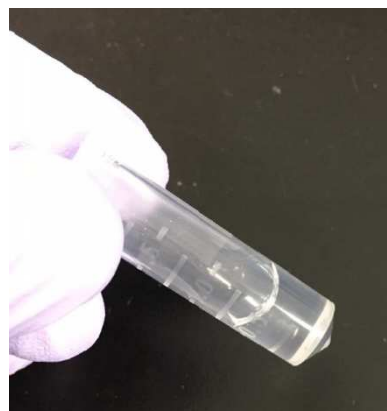
- cellnest, recombinant peptide based on human collagen type I lyophilized (hereinafter “cellnest”): 100 mg
- Water for injection: 2.4 mL

<Necessary tools>

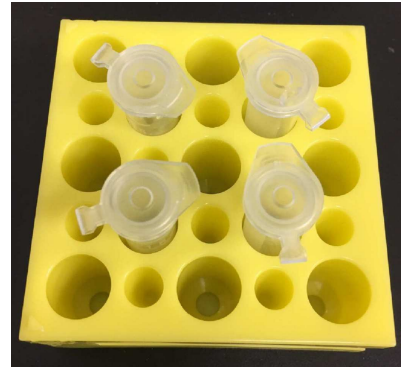
- Micro tubes (2 ml): 4
- Freezer (-20°C)
- Freeze dryer
- Vacuum oven

[Protocol]

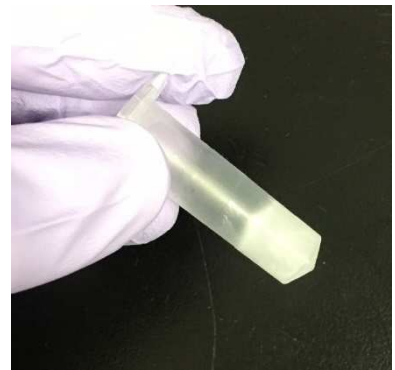
- (1) Add 2.4 mL of injection water to a 100 mg cellnest vial.
- (2) Invert the container so that the injection water reaches all of the cellnest.
- (3) Keep it for 1 hour in a 37°C incubator and let the cellnest dissolve.



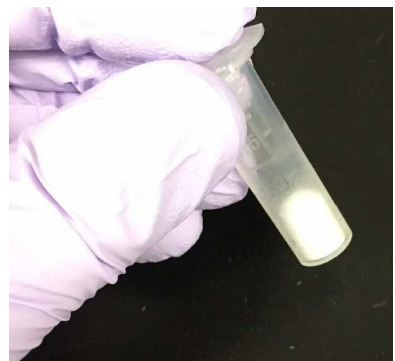
- (4) Add the cellnest solution (0.6 mL) in Step (2) into micro tubes (2 mL) and degas with centrifuge.



- (5) Keep the micro tubes overnight in a freezer (-20°C) so that the solution freezes.



- (6) Freeze-dry it in a pre-cooled freeze dryer to make the cellnest sponge (maybe good to give information on the lyophilisation process settings, e.g. plate temperature and pressure)



- (7) Take the cellnest sponge out of the micro tubes and crosslink them for 10 hours in a vacuum oven (160°C, pressure?).