FUJIFILM



Code No. 291-50502

DNA Extractor® WB Kit (Genomic DNA Extraction/Nal Method)

Obtaining super macromolecular genomic DNA from whole blood and culture cell is a primary requisite for genetic analyses of genome and molecular biology procedures. A variety of DNA purification procedures involve either hazardous processes such as the phenol/chloroform extraction or time-consuming and inefficient procedures such as lengthy dialysis and the transfer of DNA from one tube to another. The "DNA Extractor" WB Kit" employs a new extraction procedure for DNA purification after the lysis of cells. This procedure, using Sodium Iodide (NaI) as a chaotropic agent, realizes intact DNA isolation of both purity and high yield. Furthermore, extraction is done through several brief microfuge centrifugations from a single microfuge tube. This kit is for laboratory use only.

[Reagents (50 Assays)]

One kit includes;

(1)	296-50513	Lysis Solution	65 mL	2 bottles
(2)	297-50521	Enzyme Reaction Solution	10 mL	1 bottle
(3)	294-50531	Sodium Iodide Solution	15 mL	1 bottle
(4)	291-50541	Washing Solution (A)	50 mL	1 bottle
(5)	298-50551	Washing Solution (B)	50 mL	1 bottle
(6)	295-50561	Protease	10 mg	1 bottle

[Materials and Apparatus]

Reagent	1) Isopropyl alcohol	25 mL
	2) Distilled deignized water	06 mI

Apparatus 1) Microcentrifuge (Max. 12,000 g)

2) Microcentrifuge tube with screwed cap (1.5 mL)

3) Microfuge tube mixer

Note: During the storage, in such a case, Enzyme Reaction Solution and the NaI Solution may crystalize out, solubilize the precipitate completely by warming the solution at 50° C.

[Principle of DNA Extraction]

The procedure includes three major steps : 1) Isolation of nucleus fraction. 2) Liberation of DNA, 3) Purification of DNA.

In the first step, cells are lysed by a non-ionic surfactant, polyoxyethylene (10) oxyphenyl ether, followed by a brief centrifugation. By repeating this operation, the majority of cellular macromolecules, including RNAs, are removed from the nucleus fraction. In the next step, the nucleus fraction is treated with protease in the presence of 1% sodium dodecyl sulfate for 1 hour. By this treatment DNA packed in the nucleus is expanded and liberated from nucleus protection. Simultaneously, protein in the solution is digested into polypeptide. In the final step, the resultant solution is subjected to NaI extraction of DNA as

described in literature. Polypeptide and other biological molecules remain soluble in a high concentration of Nal, a chaotropic salt, even after isopropanol is added to the solution to precipitate the nucleic acid.

[Storage]

Store at $2 \sim 10^{\circ}$ C.

[Protocol]

(Preparation of Reagents)

10 mg of Protease is reconstituted by adding 0.6 mL of distilled water. Protease solution reconstituted can be stored at -20°C.

⟨Scheme I: DNA Extraction Procedure from Whole Blood⟩

- 0.5 mL of whole blood, prepared with EDTA-2Na as anti-coagulant, is dispensed into a 1.5 mL microcentrifuge tube with screwed cap and then placed standing on ice.
- 2. After adding 0.5 mL of Lysis Solution and capping the tube, mix gently several times by inversion. (Follow this for the mixing operations below.)
- 3. After a centrifugation at 10,000×g, for 20 seconds at 4°C, carefully remove the supernatant, keeping the dark red precipitate intact.
- 4. Add 1 mL of Lysis Solution to the tube and set the capped tube on a microfuge tube mixer for 30 seconds at a moderate speed.
- 5. After a centrifugation of the mixture at 10,000×g for 20 seconds at 4°C, remove the supernatant from the tube.
- 6. Repeat operation step 4-5.
- 7. Suspend the resultant pellet in $200\,\mu\text{L}$ of Enzyme Reaction Solution. Add $10\,\mu\text{L}$ of Protease Solution to the suspension and mix gently by inversion.
- 8. Incubated at 37% for 1 hour. (During the incubation, mix the solution several times by inversion.)
- After the 1 hour-incubation, add 0.3 mL of NaI Solution to the mixture and mix well by inversion.
- 10. Add 0.5 mL of isopropyl alcohol to the mixture and mix well until DNA, a whitish material, appears.
- 11. After centrifugation at 10,000×g for 10 minutes at room temperature, gently removed the resultant supernatant. Put the tube upside down to remove the solution remaining on the surface of the tube.
- 12. After adding 1 mL of Washing Solution (A), mix thoroughly so that the pellet is removed from the tube wall.
- 13. After a centrifugation of the mixture at $10,000 \times g$ for 5 minutes at room temperature, remove the supernatant from the tube.
- 14. After adding 1 mL of Washing Solution (B), mix thoroughly so that the pellet is removed from the tube wall.
- 15. After a centrifugation of the mixture at 10,000×g for 5 minutes at room temperature, remove the supernatant from the tube.
- 16. The resultant pellet is vacuum-dried for about 3 minutes. (Drying should be made within 3 minutes because DNA completely dried may be hard to be dissolved).

$\langle Scheme \, II: DNA \; Extraction \; Procedure \; from \; Culture \; Cell \; (Example: HeLa, SQ-5) \rangle$

1. Culture cell pellet $(10^3 \sim 10^6 \text{ cells})$ such as HeLa and SQ-5, collected in a 1.5 mL microfuge tube is suspended in 1 mL Lysis Solution and then set the 1.5 mL microcentrifuge tube with screwed cap in a

microfuge tube mixer.

2. After a centrifugation of the mixture at 3,000×g for 5 minutes at 4°C, remove the supernatant from the tube. Follow with operations through step 4-16 as described in "Scheme I DNA Extraction Procedure from Whole Blood".

[Application of "DNA Extractor WB Kit" for volume reduction procedure

Genomic DNA was extracted from $100 \,\mu\text{L}$ of whole blood with the "DNA Extractor WB Kit" using smaller volumes of reagents than the standard procedure. In these procedures, 1/5 or 1/4 of the original reagent volumes were employed, Microfuge tubes were 0.6 mL. As a reference, genomic DNA was extracted from 500 µL of whole blood using the standard procedure.

Recovery and purity of genomic DNA

Sample Volume	Reagents	Recovery (μ g DNA/mL whole blood)	260/280
100 μL wholeblood	1/5	23.8	1.97
100 μL wholeblood	1/4	22.2	1.98
500 μL wholeblood	1/1	22.0	1.92

Note

- 1. DNA extracted from this kit is suitable for several applications, including restriction enzyme digestion and Southern blot analysis.
- 2. Whole blood anti-coagulated by EDTA is appropriate as a sample for this kit, but DNA isolated from heparinized whole blood with greater than 20 units/mL may influence the result of some experiments.
- 3. This kit is applicable to $100 \,\mu L \sim 500 \,\mu L$ of whole blood by proportionally varying the volumes of solutions used in the standard method described below. (1 mL of whole blood can be applied by using a 2 mL microcentrifuge tube and doubling the amount of each reagent.)
- 4. Whole blood stored refrigerated for 3 weeks did not show a significant decrease in the recovery of DNA with this kit.
- 5. This kit is not compatible for DNA extraction from smear preparation samples.

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全血中DNA抽出用 DNA エキストラクター®WB キット

(よう化ナトリウム法)

【はじめに】

ヒトゲノム遺伝子を分析するためには、高分子のゲノム DNA を調製す る必要があります. 現在, 種々のゲノム DNA 抽出法が行なわれていま すが、それらの大半はフェノールなどの有機溶媒系を用いるものであ り、その有毒性と共に手間と時間を要する抽出操作を行なわなければ ならないのが欠点です.

DNA エキストラクター®WB キットは主に全血液からのゲノム DNA 抽出を対象に有機溶媒を使用することなく、一連の抽出操作を一本の マイクロチューブ内で行なう様に簡素化されたものです.

容】 (50 回用: サンプル 500 µ L)

(1)	296-50513	溶解液	65mL	2本
(2)	297-50521	酵素反応液	10mL	1本
(3)	294-50531	よう化ナトリウム溶液	15mL	1本
(4)	291-50541	洗浄液 (A)	50mL	1本
(5)	298-50551	洗浄液 (B)	50mL	1本
(6)	295-50561	タンパク分解酵素	10mg	1本

(キット以外に用意するもの)

1) イソプロピルアルコール (和光特級) 25mL2) 滅菌蒸留水 0.6 mL

- 1) マイクロ遠心機
- 2) マイクロ遠心管 (1.5mL 容, スクリューキャップ)
- 3) マイクロチューブミキサー MT-360 (トミー精工) タイプ

注意:保存中に酵素反応液あるいは、よう化ナトリウム溶液に結晶が 析出した時は、50℃に加温して完全に溶かしてから使用して下 さい.

【DNA 抽出の原理】

溶解液で試料中の細胞から細胞核を取り出した後タンパク分解酵素で 核膜及び核タンパクを破壊します。そこへよう化ナトリウム溶液を添 加する事により、タンパク質及び脂質等を可溶化状態にした後、イソ プロピルアルコールで DNA を沈殿させ,回収します.

【使用法】

〈試液の調製〉

タンパク分解酵素 10mg(チューブ1本)に滅菌蒸留水を 0.6mL を加え て, 良く溶かします. (-20℃保存)

A:〈血液からの DNA 抽出〉

- 1. 全血 0.5mL (EDTA-2Na 抗凝固剤使用) を 1.5mL 容のマイクロ遠 心管に入れます.
- 2. 溶解液を 0.5mL 加えて、チューブを数回軽く、逆さにして液を混

ぜます. (下記の混合操作はこれに従って下さい.)

- 3. 遠心 (10,000×g, 20 秒, 4℃) した後, 黒っぽいペレットが流出 しないように上清を除きます.
- 4. 溶解液を 1mL 加えて、マイクロチューブミキサーにて攪拌します. (MT-360 では、スピード 7、30 秒.)
- 5. 遠心 (10,000×g, 20秒, 4℃) した後, 上清を除きます.
- 6. ステップ $4 \sim 5$ を 1 回繰り返します.
- 7. ペレットに酵素反応液 $200 \mu L$ とタンパク分解酵素 $10 \mu L$ を加えて混合します.
- 8. 37℃で1時間保温します. (途中2~3回軽く振りまぜて下さい.)
- 9. よう化ナトリウム溶液を 300 μL 加えて混合します.
- 10. イソプロピルアルコールを 0.5mL を加えて、白い綿状の DNA が 完全に見えてくるまで、混合します.
- 11. 遠心(10,000×g, 10分, 室温)した後,上清をゆっくり除きます。 容器を濾紙の上に逆さに置く等の方法で,器壁に残った溶液を充分に除いて下さい.
- 12. 洗浄液 (A) を 1mL 加えて混合します. 沈殿が器壁から剥がれる程度に充分混合して下さい.
- 13. 遠心 (10,000×g, 5分間, 室温) 後上清を除きます.
- 14. 洗浄液(B)を1mL加えて、混合します. 沈殿が器壁から剥がれる程度に充分混合して下さい.
- 15. 遠心 (10,000×g, 5分間, 室温) 後上清を除きます.
- 16. DNA 沈殿を軽く減圧乾燥します.(乾燥しすぎると, DNA が溶けにくいので,乾燥時間は,3分間以内にして下さい.)
- B:〈培養細胞(例: HeLa, SQ-5)からの DNA 抽出法〉
- 1. 培養した細胞 (例: HeLa, SQ-5) を PBS 等で $10^3 \sim 10^6 Cells/mL$ に調整し、被験液とし、1.5mL 容マイクロ遠心管に入れます.
- 遠心分離(3,000×g, 5分, 4℃)後上清を除去します。以下、A: 〈血液からの DNA 抽出法〉4~16 に準ずる。

【小スケールサンプルの応用】

ヒト全血 100μ L からゲノム DNA を抽出する場合は 0.6mL のマイクロチューブを用い、試薬は 1/4 又は 1/5 の容量で行なって下さい. DNA の回収は全血 500μ L の時とほとんど変わりません.

1. DNA の回収量及び純度(表)

サンプル量	試薬量	回収量(μg DNA/mL 全血)	260/280
100 µL 全血	1/5	23.8	1.97
100 µL 全血	1/4	22.2	1.98
500 µL 全血	1/1	22.0	1.92

- 注意:① この方法で抽出した DNA 標品には RNA が多少混在しますが、遺伝子増幅法や制限酵素処理などには、影響ありません.
 - ② 採血の抗凝固剤には、EDTA を使用して下さい. ヘパリンを 20units/mL 以上の濃度で使用した血液から単離した DNA は 制限酵素解析やその他の分析等に適さない場合があります.
 - ③ 本キットは 100μ L $\sim 500\mu$ L の全血から DNA を抽出する事が出来ます. (また、2mL 用のマイクロチューブを用い、添加する試薬をそれぞれ 2 倍にして使用すれば 1mL の全血まで可能です。)
 - ④ 約3週間まで冷蔵保存した血液サンプルからの DNA 回収率 には著しい低下は見られません.
 - ⑤ 本キットは、塗抹標本等のサンプルからの DNA 抽出には使 えません.

【貯 法】 2 ~ 10℃

【**包 装**】 50 回用(サンプル 500 µL)

製造発売元

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