FUJIFILM Wako Pure Chemical Corporation
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Code No. 012-28521 (20 µL)
018-28523 (100 µL)

Anti Iba1, Rabbit Monoclonal Antibody (6A4), recombinant

Code No.	Product name	Pkg. size	Storage
012-28521	Anti Iba1, Rabbit Monoclonal Antibody (6A4), recombinant	20 µL	-20°C
018-28523		100 µL	

Product Summary

Reactivity	Mouse, rat (not studied in other species)
Clonality	Monoclonal (Clone No. 6A4)
Host	Rabbit
Conjugate	Unconjugated
Concentration	Described on the label

Product Details

Source	CHO-Spica cell derived antibody
Immunogen	Synthetic peptide corresponding to the C-terminus of Iba1
Purity	Protein A affinity chromatography
Isotype	IgG
Form	Liquid
Preservative	0.05% Sodium azide
Buffer	PBS with 50% glycerol

Application

Immunohistochemistry (frozen section) 1:200 - 10,000

* The optimal dilutions/concentrations should be determined by each laboratory.

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Value from Innovation

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Recommend Protocol

Step 1. Tissue preparation

1. The animal (mouse or rat) should be fixed by perfusing with 4% paraformaldehyde-PBS under the deep anesthesia.
2. After dissection the tissue, sink the tissue in 4% paraformaldehyde-PBS for post-fixation, following to 30% sucrose/ 4% paraformaldehyde-PBS, respectively.
   ∗Sink the tissue until falling-down on the bottom of tube.
3. Mount the tissue with sectioning compound and freeze it completely.
4. Prepare a 20-50 μm thick of frozen section using the cryostat and mount the slice on the pre-coated slide glass for IHC.

Step 2. Blocking and Permeabilization

1. Rinse the section several times with PBS for removing the sectioning compound.
2. Block the section by Blocking solution (1% animal serum, 2% BSA and 0.3% TritonX-100 in PBS) for 2 hours at room temperature.

Step 3. Antibody staining

1. Incubate the section with primary antibody solution (Anti Iba1, Rabbit Monoclonal Antibody (6A4), recombinant, 1000 times dilution in Blocking solution) at 4℃ over night.
2. Wash the section three times with 0.3% TritonX-100 in PBS for 5 min.
3. Incubate the section with secondary antibody solution (i.e. fluorescent dye-conjugated anti-rabbit IgG, appropriatedilution in Blocking solution) for 2 hours RT.
4. Wash the section three times with 0.3% TritonX-100 in PBS for 5 min.

Optional: Double or nuclear staining

(a) Double staining

· Prepare the different primary and secondary antibodies than those used in “Step 3” and repeat “Step 3”.
  ∗You must prepare a source of primary antibody that is different from the one used in “Antibody staining”.

(b) Nuclear stain

· Prepare DNA binding dye such as DAPI (i.e. code No. 342-07431). The staining protocol follows instruction manual of each dye.

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Step 4. Mounting and Detection

1. Rinse the section with ddH2O quickly, and snap-off the slide.
2. Absorb remaining ddH2O with paper and drop anti-fade mounding medium, then place a coverslip on the section.
3. Observe the section image by fluorescent microscopy or confocal microscopy with appropriate filter set.