A novel affinity-based method for the isolation of highly purified extracellular vesicles

Masayuki Yamane⁴, Wataru Nakai¹, Takeshi Yoshida^{1,2}, Diego Diez³, Yuji Miyatake^{1,2}, Takahiro Nishibu⁴, Ryo Ukekawa⁴, Naoko Imawaka⁴, Ken Naruse⁴, Yoshifusa Sadamura⁴ & Rikinari Hanayama^{1,2,5}

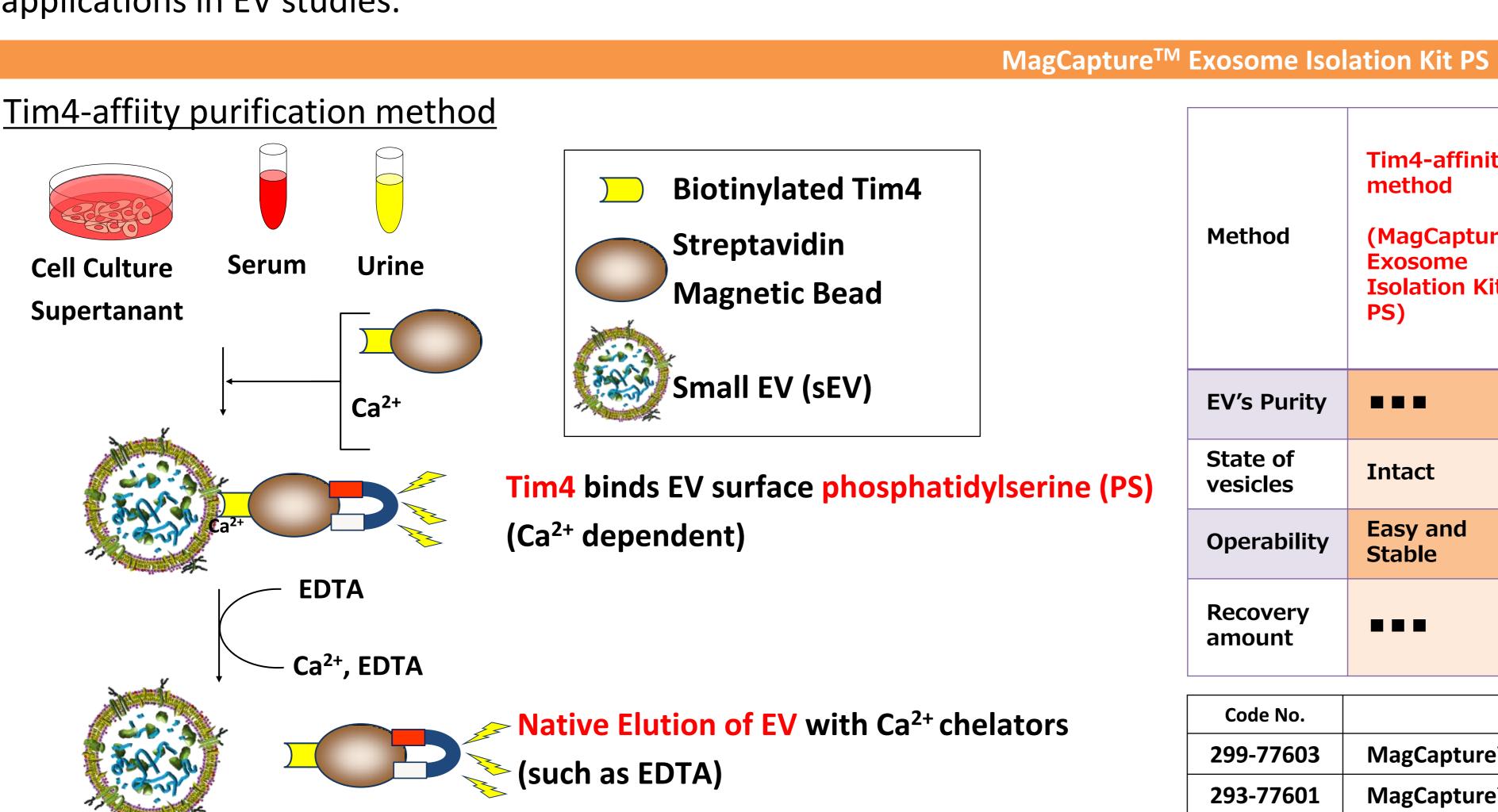
- 1 Laboratory of Immune Network, WPI Immunology Frontier Research Center (IFReC), Osaka University, Japan
- 2 Department of Immunology, Kanazawa University Graduate School of Medical Sciences, Japan
- 3 Quantitative Immunology Research Unit, WPI Immunology Frontier Research Center (IFReC), Osaka University, Japan
- 4 Wako Pure Chemical Industries Ltd, Japan
- 5 PRESTO, Japan Science and Technology Agency (JST), Japan

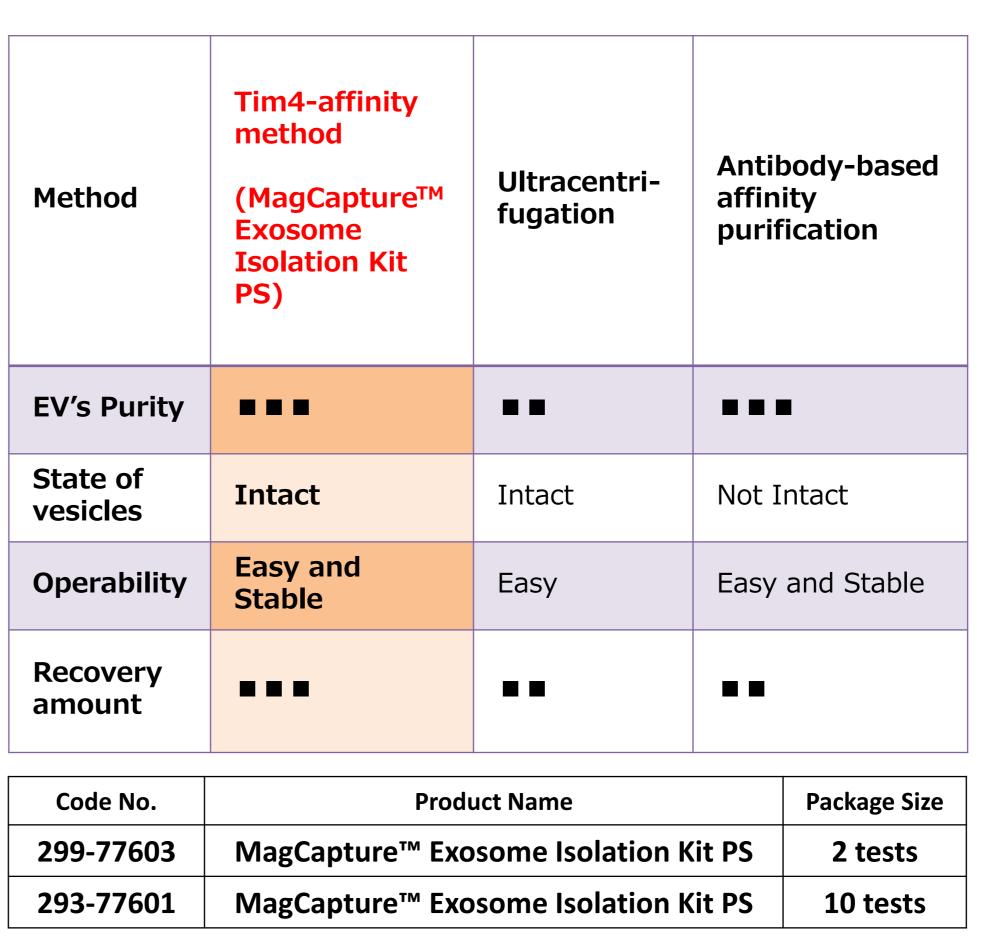




<u>Abstract</u>

Extracellular vesicles (EVs) such as exosomes and microvesicles serve as messengers of intercellular network, allowing exchange of cellular components between cells. EVs carry lipids, proteins, and nucleic acids derived from their producing cells, and have potential as biomarkers specific to cell types and even cellular states. However, conventional methods (such as ultracentrifugation or polymeric precipitation) for isolating EVs have disadvantages regarding purity and feasibility. Here, we have developed a novel method for EV purification by using Tim4 protein, which specifically binds the phosphatidylserine displayed on the surface of EVs. Because the binding is Ca²⁺-dependent, intact EVs can be easily released from Tim4 by adding Ca²⁺ chelators. Tim4 purification, which we have applied to cell conditioned media and biofluids, is capable of yielding EVs of a higher purity than those obtained using conventional methods. Tim4 protein can also be used as a powerful tool for quantification of EVs in both ELISA and flow cytometry formats. Therefore, the affinity of Tim4 for EVs will find abundant applications in EV studies.





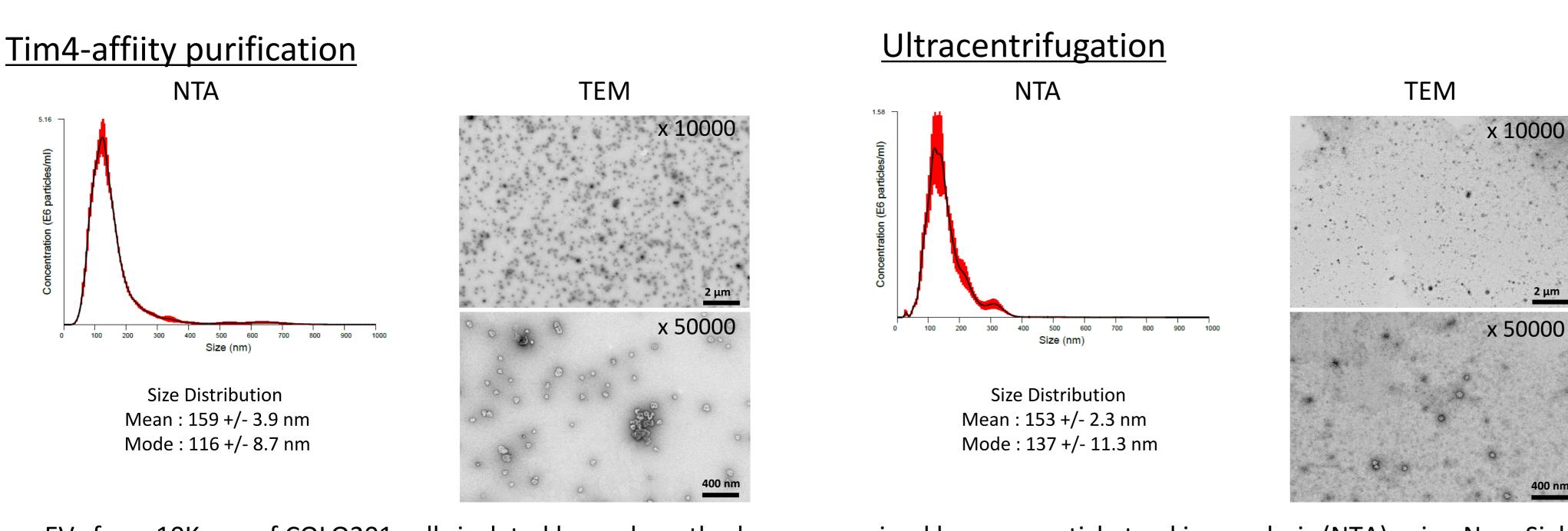
MagCpatureTM Exosome Isolation Kit PS



Sample Type: cell culture supernatant, serum, plasma, urine, etc.

◆ MagCpatureTM Exosome Isolation Kit PS can purify any EVs which expose phosphatidylserine on the outer surface of their lipid bilayer. It has been confirmed that human, mouse, and bovine EVs can be purified by this isolation kit.

Particle analysis of sEVs purifeid by the Tim4-affinity methods



sEVs from 10K sup of COLO201 cells isolated by each method were examined by nanoparticle tracking analysis (NTA) using NanoSight and transmission electron microscope (TEM)

The appearance of sEVs isolated by the Tim4affinity purification method matched the
typical saucer-like shape as previous reported*,
and almost no contaminants could be observed.
In contrast, sEVs isolated by UC were
accompanied by a large number of small
precipitates probably derived from
supplements added in advance into the
medium.

*Raposp, G. *et al.* (1996)

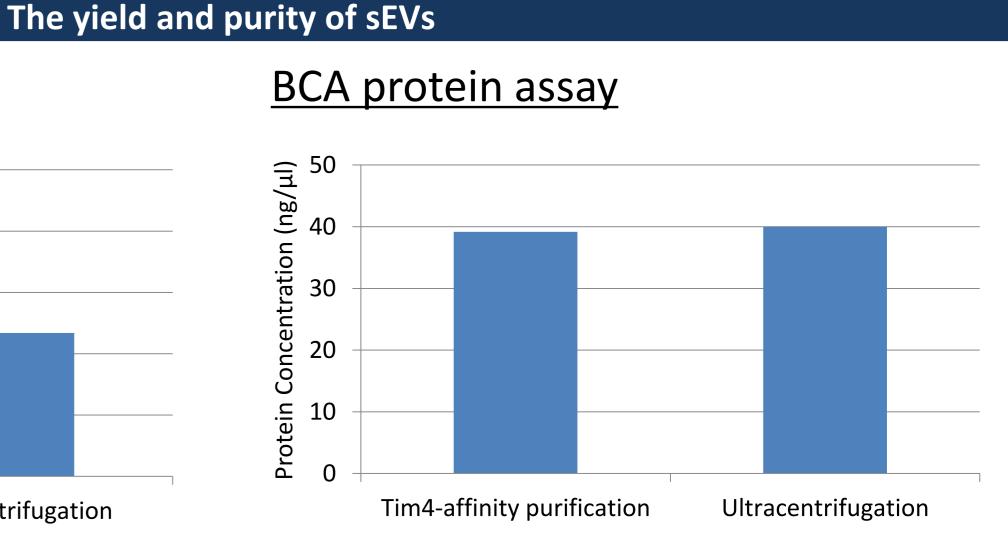
PS CaptureTM Exosome ELISA Kit Tim4-based ELISA method Substrate Samples: Purified sEVs from COLO201 cell culture supernatant Abs. 450 nm (sEVs were purified by the MagCaptureTM Exosome Isolation Kit PS) Western blotting **POD-labeled Anti-mouse IgG** sEV Surface Antigen Limit of Detection was about **75 ng or 0.5ng** im4-based ELISA Tim4-immobilized 96-well plate Standard curve of COLO201 2.0000 Limit of Limit of 1. sEV binds to the Tim4 immobilized on the 96-well plate quantification **ב** 1.5000 detection (Blank+10SD) (Blank+ 2. Detect the sEV with sEV surface antigen antibody 1.0000 3.3SD) ₹ 0.5000 and POD-labeled Anti-Mouse Antibody 0.11 ng/mL 0.34 ng/mL (Total 11 pg) (Total 34 pg) Purifed exosome (ng/mL Code No. **Product Name Package Size** The sensitivity of the Tim4-based ELISA was 50 to 1,000 times PS CaptureTM Exosome ELISA Kit 297-79201 higher than that of western blotting. 96 Reactions

(Anti-mouse IgG POD)

that by the ultracentrifugation.

Tim4-affinity purification

2.5
(weight 2.0
2.5
1.5
0.0
Tim4-affinity purification
Ultracentrifugation



sEVs from 10K sup of COLO201 cells isolated by each method were examined by Tim4-based ELISA analysis and BCA protein assay

The Tim4-affinity purification method could isolate about twice as many sEVs as the Ultracentrifugation method. In contrast, equal amount of proteins were detected in sEV fractions using BCA protein assay.

The Uptake of sEV **Lipid staining** by PKH67 Ultracentrifugation Tim4-affinity Ultracentrifugation Negative Control $(5\mu g, 2.8x10^{10} particles sEVs)$ (2.8x10¹⁰ particles sEVs) (5µg sEVs) Tim4-affinity (5μg, 2.8x10¹⁰ particles) Ultracentrifugation (5μg, 3.4x10¹⁰ particles) **Phase** Ultracentrifugation (4.1μg, 2.8x10¹⁰ particles) Signal Intensity RNA staining by SYTO RNASelect Negative Control Hoechst33342 **PKH67 5μg, 1.8x10¹⁰ particles**) **Ultracentrifugation** (5μg, 1.6x10¹⁰ particles) (5.6µg, 1.8x10¹⁰ particles) Signal Intensity

Microarray analysis of miRNA $\frac{1}{5000} = \frac{1}{5000} = \frac{1}{5000}$

miRNA microarray analysis of sEVs from COLO201 cells was performed by the 3D-Gene (TORAY)

miRNA microarray analysis of sEVs from COLO201 cells revealed high correlation of miRNA profiles between Tim4-affinity purification method and ultracentrifugation method.

Conclusions

• Tim4-affinity purification method could be used for the efficient isolation of sEVs

sEVs from COLO201 cells were labeled by the fluorescence dye and the uptake of these sEVs by HeLa cells was examined.

sEVs purified by the Tim4-affinity purification method were more efficiently incorporated into the HeLa cells than

- Tim4-affinity purification method can purify intact extracellular vesicles.
- sEVs purified by the Tim4-affinity purification method were efficiently taken up by the recipient cells.
- High correlation of microRNA profiles between Tim4-affinity purification method and ultracentrifugation was revealed.