

## TECHNICAL DATA SHEET

# COLUMBIA AGAR (BASE)

## CULTURE & ISOLATION OF FASTIDIOUS MICROORGANISMS

### 1 INTENDED USE

Columbia Agar is a highly nutritive medium used for the growth and isolation of a large variety of microorganisms, particularly very fastidious bacteria : streptococci and pneumococci in animal samples. When blood, selective agents or growth accelerators are added, it becomes possible to prepare a wide variety of media adapted to specific uses.

### 2 HISTORY

Developed by Ellner in 1966, Columbia Agar enables luxuriant colonies, perfectly defined hemolytic zones and well characterized colonies and pigmentation to be obtained.

### 3 PRINCIPLES

Peptones included in the composition of the medium favor the excellent growth of colonies.

Yeast extract is a source of vitamin B complex.

Starch is a detoxifying agent and also an energy source.

Defibrinated sheep blood, which can be added to the medium, favors the detection of hemolytic reactions and supplies X factor (heme) required for the growth of a large number of bacteria, but lacks V factor (nicotinamide adenine dinucleotide) due to the presence of an NADase, which destroys any NAD present. *Haemophilus influenzae*, which requires both X and V factors, does not grow on agar containing ordinary blood.

The following media can be prepared with Columbia base :

- **Blood agar** : By adding 5 or 10% sterile sheep blood after autoclaving and cooling, the medium is suitable for the growth of *Streptococcus*, *Pneumococcus*, *Staphylococcus*, *Listeria* and *Erysipelothrix*. It can be made selective by adding colistin and nalidixic acid to preclude the development of Gram-negative bacteria and *Bacillus*.
- **Chocolate agar** : By adding 10% sheep or horse blood to sterile Colombia Agar and heating to 70°C for 5 minutes until a chocolate color develops, an excellent medium is obtained for growth of *Haemophilus*, *Neisseria*, *Taylorella* or *Campylobacter*.
- **Base media without enrichment** : Columbia Agar can be used to grow *Brucella abortus*, *Yersinia pestis* and *Clostridium perfringens*, as well as all enterobacteria.

### 4 TYPICAL COMPOSITION

The composition can be adjusted in order to obtain optimal performance.

For 1 liter of base media :

- Polypeptone .....	23,0 g
- Starch .....	1,0 g
- Sodium chloride .....	5,0 g
- Bacteriological agar.....	13,5 g

pH of ready-to-use media at 25 °C : 7,3 ± 0,2.

## 5 PREPARATION

- Suspend 42.5 g of dehydrated medium (BK019) in 1 liter of distilled or deionized water.
- Slowly bring to boiling, stirring with constant agitation until complete dissolution.
- Dispense in tubes or flasks.
- Sterilize in an autoclave at 121°C for 15 minutes.
- Cool and maintain at 44-47°C.
- Aseptically add 5 to 7 mL of sterile, defibrinated sheep blood per vial.
- Mix well.
- Pour into sterile Petri dishes and let solidify on a cool surface.
- Dry the plates with the covers partially removed.

✓ **Reconstitution :**

42,5 g/L

✓ **Sterilization :**

15 min at 121°C

### NOTE :

For other applications, use the corresponding protocol.

## 6 INSTRUCTIONS FOR USE

- Inoculate in order to obtain isolated colonies.
- Incubate at 37 °C for 24 to 48 hours in optimal conditions for the culture of the inoculated germs.

## 7 RESULTS

Observe the bacterial growth.

### Beta hemolysis

*Streptococci* belonging to Lancefield group A appear as small, grey colonies, translucent or opaque, surrounded by a zone of beta hemolysis. Other bacteria may present the same type of hemolysis : *Listeria*, hemolytic *Staphylococci*, *Escherichia coli* and *Pseudomonas*.

*Staphylococci* appear as opaque, yellow-gold or white colonies, with or without type β hemolysis zones.

*Listeria* present small zones of beta hemolysis.

*Bacillus cereus* form a clear zone surrounding the colonies.

See ANNEX 1 : PHOTO SUPPORT.

### Alpha hemolysis

*Pneumococci* appear as flat, shiny, grey and occasionally mucoid colonies surrounded by a zone of narrow, greenish hemolysis referred to as alpha hemolysis.

### CAMP Factor

Group B *Streptococci* produce an extracellular, thermostable substance (CAMP Factor) which provokes a triangle of total hemolysis in a zone of incomplete staphylococcal hemolysis, at the junction of the two cultures.

## 8 QUALITY CONTROL

**Dehydrated media** : beige powder, free-flowing and homogeneous.

**Prepared media** : (with 5% defibrinated sheep blood) : opaque, red agar.

Typical culture response after 48 hours of incubation at 37 °C, with 5% sheep blood (qualitative method of inoculation):

Microorganisms	Growth	Type of hemolysis
<i>Streptococcus pyogenes</i>	ATCC® 19615	Good, score 2
<i>Streptococcus pneumoniae</i>	ATCC 6303	Good, score 2
<i>Listeria monocytogenes</i>	ATCC 19115	Good, score 2
<i>Staphylococcus aureus</i>	WDCM 00034	Good, score 2
<i>Escherichia coli</i>	WDCM 00013	Good, score 2

## **9      STORAGE / SHELF LIFE**

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**Dehydrated base media :** 2-30 °C.

The expiration date is indicated on the label.

**Prepared based media in vials (\*) :** 180 days at 2-8 °C.

**Prepared base media with sheep blood (\*) :** 30 days at 2-8 °C.

(\*) Benchmark value, determined in standard conditions of preparation, following manufacturer's instructions.

## **10     PACKAGING**

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**Dehydrated media :**

500 g bottle ..... BK019HA

## **11     BIBLIOGRAPHY**

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Neter, E.. 1947. The effect of yeast concentrate on the growth and survival of *Haemophilus influenza* in infusion broth. Journal of Bacteriology, **54** : 70-71.

Thayer, J.D. and Martin, J.E.. 1966. Improved medium selective for cultivation of *Neisseria gonorrhoeae* and *Neisseria meningitidis*. Public Health Report, **81** : 559-562.

Ellner, P.D., Stoessel, C.J., Drakeford, E. and Vasi, F.. 1966. A new culture medium for medical bacteriology. American Journal of Clinical Pathology, **45** : 502-504.

NF V08-405. Décembre 1986. Conserves. Recherche des *Clostridium* thermophiles.

NF U47-108. Décembre 2012. Méthodes d'analyse en santé animale. Isolement et identification de *Taylorella equigenitalis* à partir de prélèvements génitaux d'équidés.

NF EN ISO 10272-1. Avril 2006. Microbiologie des aliments. Méthode horizontale pour la recherche et le dénombrement des *Campylobacter* spp. Partie 1 : Méthode de recherche.

NF EN ISO 10272-1. Juillet 2017. Microbiologie de la chaîne alimentaire - Méthode horizontale pour la recherche et le dénombrement de *Campylobacter* spp. - Partie 1 : méthode de recherche.

NF EN ISO 10272-2. Juillet 2017. Microbiologie de la chaîne alimentaire - Méthode horizontale pour la recherche et le dénombrement de *Campylobacter* spp. - Partie 2 : technique par comptage des colonies.

Pharmacopée Européenne. Chapitre 2.6.13. Contrôle microbiologique des produits non stériles : Recherche de microorganismes spécifiés.

## **12     ADDITIONAL INFORMATION**

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The information provided on the labels take precedence over the formulations or instructions described in this document and are susceptible to modification at any time, without warning.

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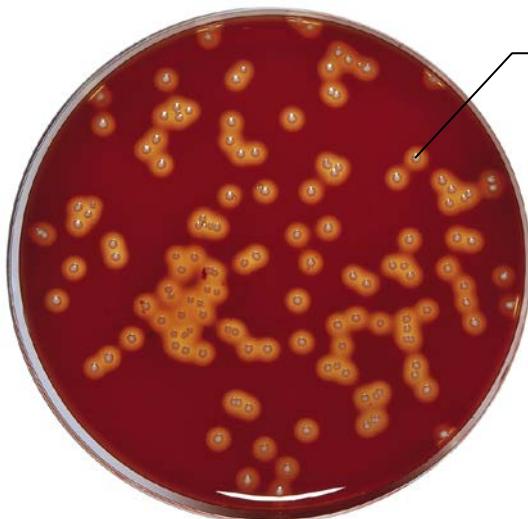
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### **Columbia Agar (base)**

A highly nutritive media allowing the culture and isolation of a large variety of microorganisms.

#### **Results :**

Agar with 10% sterile sheep blood added.  
Incubation 48 hours at 37 °C.



#### **Group D Streptococci**

Characteristic colonies surrounded by a zone of clear hemolysis ( $\beta$ -hemolysis)