

Code No. 294-85701

EV-Perm™ Permeabilization Pretreatment Kit for Exosome Membrane

[Product Information]

Code No.	Product Name	Volume	Storage
294-85701	EV-PermTM Permeabilization Pretreatment Kit for Exosome Membrane	1 kit	Frozen (-20℃)

[Kit Contents]

Reagent	Volume	Storage
Reagent A (20×)	500 μL×1	Frozen (-20℃)
Doggont P. (10v)	E ml v 1	Before thawing : -20℃
Reagent B (10×)	5 mL×1	After thawing : 2~10℃
Descent C (100y)	F00l x 1	Before thawing : -20℃
Reagent C (100×)	500 μL×1	After thawing : 2~10℃

[How to store each reagent]

Store each reagent according to the storage condition above.

Droparation	of reagents	
Preparation	or readents	

1. Reagent A (20×)

Following thaw, mix the vial by gently tapping, and prepare aliquots. Close the lid tightly and immediately store the remaining solution in the freezer (-20° C).

- If stored in a liquid state for a long time, components may precipitate.
- If precipitation is observed, heat it at 60° C for about 30 minutes to dissolve it, then store it in the freezer.

2. Reagent B (10 \times) and Reagent C (100 \times)

Following thaw, mix the vial by gently tapping, and prepare aliquots. Close the lid tightly and immediately store the remaining solution in the refrigerator ($2\sim10^{\circ}$ C).

[About this product]

This product is a reagent that enables the detection of internal markers of extracellular vesicles. Until now, the detection targets of the PS CaptureTM Exosome ELISA Kit Series and PS CaptureTM Exosome Flow Cytometry Kit were limited to the detection of surface markers of extracellular vesicles. However, when used in combination with this product, it is possible to improve the permeability of the extracellular vesicle membrane surface and detect internal markers.

For PS Capture[™] Exosome ELISA Kit

[Equipment a	nd reagents to be supplied by user]
① ELISA Kit([□ Check):
□ One of th	e following kits
Code No.	Product Name
297-79201	PS Capture™ Exosome ELISA Kit (Anti Mouse IgG POD)
298-80601	PS Capture™ Exosome ELISA Kit (Streptavidin HRP)
② Reagents (☐ Check):
□ Distilled W	/ater
☐ Specific ar	ntibody for Extracellular vesicle internal protein
If using the	PS Capture™ Exosome ELISA Kit (anti-mouse IgG POD), please prepare a mouse
monoclonal	antibody, and if using the PS Capture TM Exosome ELISA Kit (Streptavidin HRP), prepare
a biotin-labe	eled antibody.
③ PS Capture ^{TI}	^M Exosome ELISA Kit included reagents
□ Washing E	Suffer (10×)
☐ Exosome I	Binding Enhancer (100×)
□ Reaction E	Buffer
Please use t	he Exosome Binding Enhancer and Reaction Buffer in the PS Capture™ Exosome ELISA
Kit. Even if	you use Buffer with this kit, you will still have Buffer for the PS Capture™ Exosome
ELISA Kit.	
4 Equipments	$(\Box Check)$:
□ Microwell	plate (PS Capture™ Exosome ELISA kit accessory)
□ Plate seal	(PS Capture™ Exosome ELISA kit accessory)
□ Microplate	shaker
☐ Microplate	reader

[Prepartaion of samples]

For PS Capture[™] Exosome ELISA Kit (anti-mouse IgG POD)

It is necessary to optimize the concentration of extracellular vesicles used to detect the target marker using this product. The optimal concentration of extracellular vesicles was determined using the PS Capture™ Exosome ELISA kit (anti-mouse IgG POD) such that the resulting absorbance (450 nm minus 620 nm) was in the range of 0.2 to 2.5. To adjust the concentration of extracellular vesicles, dilute them with the Reaction/Wash Buffer (1x) included in the kit.

For PS CaptureTM Exosome ELISA Kit (Streptavidin HRP)

It is necessary to optimize the concentration of extracellular vesicles used to detect the target marker using this product. The optimal concentration of extracellular vesicles was measured using the PS Capture[™] Exosome ELISA Kit (Streptavidin HRP) and adjusted so that the resulting absorbance (450 nm minus 620 nm) was in the range of 0.2 to 2.5. To adjust the concentration of extracellular vesicles, dilute at least two times with the Reaction Buffer included in the kit.

(Prepartion of reagents)

1. Preparation of permeabilization solution

Dilute each reagent, referring to Table 1 below. After dilution, mix well.

Table 1. Dilution ratio and amount of reagent added for each reagent used in permeabilization solution

Donash	Dilution	12 samples [*]	24 samples [*]	48 samples [*]
Reagent	Dilution	(24 wells)	(48 wells)	(96 wells)
Distilled Water	-	2.1 mL	4.2 mL	8.4 mL
Reagent A (20×)	1:20	125 µL	250 μL	500 μL
Reagent B (10×)	1:10	250 μL	500 μL	1.0 mL
Reagent C (100×)	1:100	25 μL	50 μL	100 µL
Total	-	2.5 mL	5.0 mL	10.0 mL

2. Preparation of negative control solution

Dilute each reagent referring to Table 2 below. After diluting, mix well (Do not add Reagent A when preparing the negative control solution.)

Table 2. Dilution ratio and amount of reagent added for each reagent used for negative control

Reagent	Dilution	12 samples [*] (24 wells)	24 samples [*] (48 wells)	48 samples [*] (96 wells)
Distilled Water	-	2.2 mL	4.5 mL	8.9 mL
Reagent B (10×)	1:10	250 µL	500 μL	1.0 mL
Reagent C (100×)	1:100	25 µL	50 μL	100 µL
Total	-	2.5 mL	5.0 mL	10.0 mL

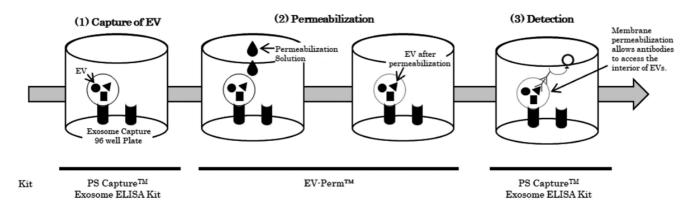
For performing in duplicate.

3. Preparation of washing solution (1x)

Washing Buffer (10x) included with the PS CaptureTM Exosome ELISA Kit was diluted 10 times with purified water. After that, add 1/100 amount of Exosome Binding Enhancer (100x) to the diluted solution and mix well. 10

[Procedure for detection using EV-Perm TM]

 \blacksquare If you use EV-Perm TM in conjunction with the PS Capture TM Exosome ELISA Kit series



[Procedure]

1. Process Flow

	Pi	rocess	Reagent · Equipment	Time	
	Cantura Evacama	Washing the plate	Washing buffer (1×)		
	Capture Exosome	Capture Exosome	Sample Capture	>2 hrs	
1.		on the plate	(1× Reaction Buffer)		
		Washing the plate	Washing buffer (1×)		
	Permeabilization	Permeabilization*	① permeabilization solution		
2.	remeabilization		② negative control solution	>1 hr	
		Washing the plate	Washing buffer (1×)		

The permeabilization solution and negative control solution are separate for each sample and detection condition in the permeabilization process.

The following steps are performed according to the PS Capture $^{\text{TM}}$ Exosome ELISA kit.

	Pi	rocess	Reagent · Equipment	Time
3.	1st antibody	1st antibody reaction	Add detection antibody	1 hr
	reaction	Washing the plate	Washing buffer (1×)	
	2nd antibody /	2nd antibody / streptavidin-HRP	2nd antibody / streptavidin-HRP	
4.	streptavidin-HRP	reaction	reaction	>1 hr
	reaction	Washing the plate	Washing buffer (1×)	
		Substrate addition	TMB Solution	
5.	Substrate	Substrate dudition	Stop Solution	>30 min
		ELISA plate reading	Microplate reader	

- 2. The procedure of #2 permeabilization in the process flow above.
- (1) Follow the instructions for the PS Capture™ Exosome ELISA kit and proceed to the washing step Procedure# 4 before adding the 1st antibody reaction solution.
- (2) Add 100 μL of permeabilization solution or 100 μL of negative control solution to the extracellular vesicle sample (well).

- (3) Apply a plate seal and incubate at room temperature for 1 hour while stirring at 500 rpm using a microplate shaker.
- (4) Decant the reaction solution and wash each well three times with 300-350 μ L of washing solution (1x).
- (5) From the process flow "3. 1st antibody reaction", proceed to step "[Procedure] 5)" according to the PS Capture™ Exosome ELISA Kit instruction manual.

Please refer to the PS Capture[™] Exosome ELISA Kit Instruction Manual for post-measurement calculations and analysis methods.

For PS Capture[™] Exosome Flow Cytometry Kit

(E	quipment an	d reagents to be supplied by user]
1	PS CaptureTi	M Exosome Flow Cytometry Kit(□ Check):
	The Kit	
	Code No.	Product Name
2	297-79701	PS Capture™ Exosome Flow Cytometry Kit
2	Reagents (Check):
	Distilled Wa	ater
	Specific an	tibody for Extracellular vesicle internal protein
I	Please prepa	re fluorescent labeled antibodies.
3 1	PS Capture™	Exosome Flow Cytometry Kit included reagents
	Washing B	uffer (10×)
	Exosome B	inding Enhancer (100×)
	Exosome M	lagnetic Beads
ı	Please use th	e Exosome Binding Enhancer and Reaction Buffer in the PS CaptureT ^M Exosome Flow
(Cytometry Ki	t. Even if you use Buffer with this kit, you will still have Buffer for the PS Capture™
	Exosome Flo	w Cytometry Kit.
4	Equipments	(□ Check):
	Centrifuge	tubes (15 mL)
	Centrifuge	tubes (1.5 mL)
	Vortex Mix	er
	Desktop Ce	entrifuge
	Magnetic s	tand
	Flow Cyton	netry

[Prepartion of reagents]

1. Preparation of permeabilization solution

Dilute each reagent, referring to Table 1 below. After dilution, mix well.

Table 1. Dilution ratio and amount of reagent added for each reagent used in permeabilization solution

Doggont	Dilution	2 Reactions	4 Reactions	8 Reactions
Reagent	Dilution	(100 µL) *	(167 μ L) *	(300µL) *
Distilled Water	-	126 µL	184.8 μL	294 μL
Reagent A (20×)	1:20	7.5 µL	11 µL	17.5 μL
Reagent B (10×)	1:10	15 µL	22 µL	35 μL
Reagent C (100×)	1:100	1.5 µL	2.2 μL	3.5 µL
Total	-	150 µL	220 µL	350 μL

[%] For the amount to add, please refer to Table 2, "Sample (μ L)" in the instruction manual of the PS CaptureTM Exosome Flow Cytometry Kit.

2. Preparation of negative control solution

Dilute each reagent referring to Table 2 below. After diluting, mix well (Do not add Reagent A when preparing the negative control solution.)

Table 2. Dilution ratio and amount of reagent added for each reagent used for negative control*

Reagent	Dilution	2 Reactions	4 Reactions	8 Reactions
Reagent		(100 µL) *	(167 μL) [*]	(300µL) *
Distilled Water	-	133.5 µL	195.8 µL	311.5 μL
Reagent B (10×)	1:10	15 µL	22 µL	35 µL
Reagent C (100×)	1:100	1.5 µL	2.2 μL	3.5 µL
Total	-	150 µL	220 µL	350 μL

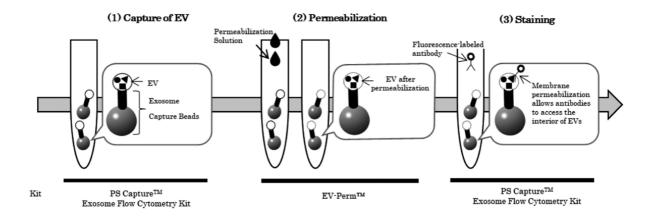
[%] For the amount to add, please refer to Table 2, "Sample (μ L)" in the instruction manual of the PS CaptureTM Exosome Flow Cytometry Kit.

3. Preparation of WB (+Enhancer)

Add 0.5 mL of Washing Buffer (10X) and 4.5 mL of purified water to a 15 mL centrifuge tube and mix by vortexing. Add 50 μ L (1/100 volume) of Exosome Binding Enhancer (100x) and mix on a vortex mixer.

This is the liquid volume for a reaction system (2 reactions) using a 1.5 mL microtube. If you wish to increase the reaction scale, please scale up according to Table 2 in the instruction manual included with the PS CaptureTM Exosome Flow Cytometry Kit.

- 4. Preparation of extracellular vesicle-bound magnetic beads
 - Prepare according to "2. Isolation of extracellular vesicles" in the instruction manual included with the PS CaptureTM Exosome Flow Cytometry Kit.
 - \blacksquare If you use EV-Perm TM in conjunction with the PS Capture TM Flow Cytometry Kit



[Procedure]

1. Process Flow

	Pr	rocess	Reagent · Equipment	Time
		Fyggang Including	WB (+Enhancer)	
1.	Exosome Isolation	Exosome Isolation	Exosome Capture Beads	>70 mins
		Capture Exosome	WB (+Enhancer)	
2.	Permeabilization	Permeabilization*	Permeabilization solution Negative control solution	>1 hr
		Washing	WB (+Enhancer)	

The following steps are performed according to the PS Capture™ Flow Cytometry kit.

Process			Reagent · Equipment	Time
3.	Antibody incubation	Antibody incubation	fluorophore-conjugated antibody or fluorophore-conjugated isotype control	>1 hr
		Washing	WB (+Enhancer)	
4.	Detection	Detection	Flow Cytometry	

- The permeabilization solution and negative control solution are separate for each sample and detection condition in the permeabilization process.
- 2. Protocol (process flow "2. Permeabilization" when analyzing one type of extracellular vesicle marker)
- (1) Follow the instructions for the PS Capture™ Exosome Flow Cytometry Kit and proceed to "2. Isolation of extracellular vesicles" to prepare extracellular vesicle magnetic beads (700 µL) for 6 reactions.
- (2) Dispense 300 µL of extracellular vesicle-bound magnetic beads into two microtubes.
- (3) Centrifuge the microtube and spin down the magnetic beads.
- (4) Leave the microtube on the magnetic stand for 1 minute, and remove the supernatant with a pipette.
- (5) Add 300 µL of WB (+Enhancer) to the microtube and mix by vortexing.
- (6) Repeat steps (3) to (5) twice.
- (7) Add 100 μ L of permeabilization solution or 100 μ L of negative control solution to each tube and mix by vortexing.
- (8) Leave the microtube from (7) at room temperature, mix by vortexing after 20 and 40 minutes, and permeabilize the exosomes for 1 hour.
- (9) Leave the microtube on the magnetic stand for 1 minute, and remove the supernatant with a pipette.
- (10) Add 300 µL of WB (+Enhancer) to the microtube and wash by vortexing.
- (11) Repeat washing steps (10) to (11) twice.
- (12) Add 300 μ L of WB (+Enhancer) to the microtube and mix by vortexing.
- (13) From the process flow "3. Staining of extracellular vesicles", proceed to step "3. Staining of extracellular vesicles" according to the PS Capture™ Exosome Flow Cytometry Kit instruction manual.

Please refer to the PS Capture[™] Flow Cytometry Kit Instruction Manual for post-measurement calculations and analysis methods.

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